

STRUCTURE OF ASZONALENIN, A NEW METABOLITE OF ASPERGILLUS ZONATUS

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Summary : Aszonalenin (1) has been isolated from Aspergillus zonatus together with LL-S490 β (2) and the structure of 1 has been established by spectroscopic evidences and chemical transformation.

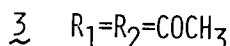
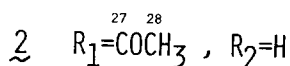
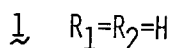
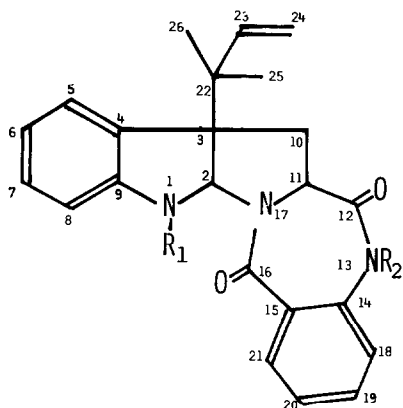
In the course of our screening for biologically active substances among fungal metabolites, we isolated a new compound from Aspergillus zonatus IFO 8817. We wish to report the isolation and structure of this compound (1) designated aszonalenin. In addition to 1, we isolated LL-S490 β (2) which had been obtained from an unidentified Aspergillus species by Ellestad et al.¹⁾

The fungus was stationarily cultured at 24°C for 21 days in the malt extract medium. The acetone extract from the dry mycelial mats was chromatographed on a column of silica gel and eluted with benzene-acetone (19:1, v/v). After elution of LL-S490 β (2), aszonalenin (1) was obtained as a crude substance, which was rechromatographed over silica gel and eluted with benzene-ethyl acetate (19:1, v/v) to give colorless crystals in a yield of 51.2 mg/10 g dry mycelia.

Aszonalenin (1), C₂₂H₂₃N₃O₂ (EI-MS, m/z 373 M⁺ and elementary analysis²⁾) mp. 244-247°C, [α]_D²⁰ +53° (c=1.31, CHCl₃), forms colorless needles (from CHCl₃-MeOH). The physicochemical data of 1 are as follows. UV λ _{max}^{EtOH} nm(ϵ); 210 (44,700), 233 (sh, 25,100), 290 (5050). IR ν _{max}^{KBr} cm⁻¹; 3400 (NH), 1700 (C=O, amide), 1640 (C=O, amide), 1620 (C=C), 1578 (Ar). MS m/z; 373 (M⁺), 304 (M⁺-69, base peak), 130. ¹H-NMR spectrum among other spectra was very similar to that of 2. Though a signal at δ 2.59 due to methyl protons of acetyl group in 2 was not observed on the ¹H-NMR spectrum of 1, an additional signal at δ ca. 7.0 assignable to -NH- appeared in the ¹H-NMR spectrum of 1. In the ¹³C-NMR spectrum of 1, signals at δ 24.2 (q) and 171.7 (s) due to acetyl group of 2 were not observed as mentioned below.

The presence of a 1,1-dimethyl-2-propenyl group in 1 was proved by the ¹H-NMR spectrum (δ 1.08, 1.16, 5.04, 5.07, 6.10)³⁾ and the ¹³C-NMR spectrum (δ 22.6, 22.9, 41.7, 114.3, 144.1)⁴⁾ together with IR absorption band (1620

cm^{-1}). A peak at m/z 304 ($M^+ - 69$) in the mass spectrum supported the presence of a 1,1-dimethyl-2-propenyl ion.⁵⁾ $\underline{1}$ was positive to Ehrlich test. The prominent peak at m/z 130 was assigned to the indoline-3-methylene ion.⁶⁾ The UV spectrum ($\lambda_{\text{max}}^{\text{EtOH}}$: 210, 233, 290 nm) suggested the presence of indoline chromophore⁷⁾ together with indicating the presence of another chromophoric unit. In the $^1\text{H-NMR}$ spectrum, a signal at δ 8.77 was assigned to an amide proton by comparison with the spectrum of $\underline{2}$. From these results and the presence of eight aromatic protons between δ 6.55-7.92, another aromatic ring except for a benzene ring of indoline moiety must be present in $\underline{1}$. The IR spectrum of $\underline{1}$ showed absorption bands at 3240, 1700 and 1640 cm^{-1} , the latter two absorptions of which indicated the dipeptide system.^{7b)} The molecular formula and the above evidences led to the presence of 3,4-dihydro-4-methyl-1H-1,4-benzodiazepine-2,5-dione moiety [UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (ϵ); 215 (32,140), 291 (2180). IR $\nu_{\text{max}}^{\text{Nujol}}$ cm^{-1} ; 1698, 1639. $^1\text{H-NMR}$ (in CDCl_3) δ ; 7.0-7.9 (4 x Ar-H)].⁸⁾ From these results obtained above, the structure of $\underline{1}$ was determined as shown in Fig.



In the $^{13}\text{C-NMR}$ spectrum of $\underline{1}$, signals at δ 33.6 (t) and 57.5 (d) were assigned to C-10 and C-11 from the multiplicities and by comparison with the chemical shifts of echinulin (lit.,⁴⁾ 29.0 and 51.6). Also, two characteristic signals at δ 170.2 and 168.0 could be easily assigned to amide carbons of C-12 and C-16, respectively.⁴⁾ As mentioned above, signals due to the 1,1-dimethyl-2-propenyl carbons were observed. Twelve signals between δ 119.3-141.8 were assigned by comparison with the chemical shifts calculated for *o*-toluidine and *o*-tolyl-isocyanate from the equation of ^{13}C substituent effects of substituted benzene.⁹⁾ The remaining two signals at δ 82.0 and 61.1 were assigned to C-2 and C-3 from the chemical shifts and multiplicities. These signals were well coincident with those of $\underline{2}$ except for signals at δ 24.2 and 171.7 due to an acetyl group.

Table. ¹H-NMR and ¹³C-NMR Spectral Data of Aszonalenin (1) and LL-S4908 (2)^a

| Position | Group | 1 | | | 2 | | | <i>Calcd.</i> |
|----------|------------------|---------------------------|-------|---------------------------|---------------------------|-------|---------------------------|---------------------------|
| | | δ_{H} , ppm | J, Hz | δ_{C} , ppm | δ_{H} , ppm | J, Hz | δ_{C} , ppm | δ_{C} , ppm |
| 1 | NH | ca. 7.0 | - | - | - | - | - | - |
| 2 | CH | 5.57 s | - | 82.0 d | 6.10 s | - | 82.4 d | - |
| 3 | -C- | - | - | 61.1 s | - | - | 60.7 s | - |
| 4 | =C- | - | - | 131.5 s | - | - | 134.4 s | 133.8 s ^b |
| 5 | =CH | 6.55 | - | 130.9 d | 6.83 | - | 130.7 d | 130.4 d ^b |
| 6 | =CH | m | - | 125.2 d | m | - | 124.8 d | 125.6 d ^b |
| 7 | =CH | | - | 125.3 d | | - | 125.3 d | 126.8 d ^b |
| 8 | =CH | 7.92 | - | 121.3 d | 8.17 | - | 124.8 d | 124.8 d ^b |
| 9 | =C- | - | - | 135.4 s | - | - | 134.8 s | 134.9 s ^b |
| 10 | CH ₂ | 2.45 dd | 14.0 | 33.6 t | 2.42 dd | 14.0 | 30.6 t | 29.0 t ^e |
| | | | 8.0 | | | 8.0 | | |
| | | 3.46 dd | 14.0 | | 3.39 dd | 14.0 | | |
| | | | 8.5 | | | 8.5 | | |
| 11 | CH | 4.02 dd | 8.0 | 57.5 d | 3.87 dd | 8.0 | 56.9 d | 55.5 d ^e |
| | | | 8.5 | | | 8.5 | | |
| 12 | C=O | - | - | 170.2 s | - | - | 169.5 s | 168.0 s ^e |
| 13 | NH | 8.77 s | - | - | 8.62 s | - | - | - |
| 14 | =C- | - | - | 149.3 s | - | - | 141.8 s | 146.6 s ^c |
| 15 | =C- | - | - | 126.8 s | - | - | 127.4 s | 124.3 s ^c |
| 16 | C=O | - | - | 168.0 s | - | - | 167.6 s | 165.8 s ^e |
| 17 | N | - | - | - | - | - | - | - |
| 18 | =CH | 6.55 | - | 109.5 d | 6.83 | - | 119.3 d | 115.2 d ^c |
| 19 | =CH | | - | 133.0 d | | - | 132.8 d | 133.6 d ^c |
| 20 | =CH | m | - | 118.9 d | m | - | 121.1 d | 118.7 d ^c |
| 21 | =CH | 7.92 | - | 128.5 d | 8.17 | - | 129.1 d | 129.5 d ^c |
| 22 | -C- | - | - | 41.7 s | - | - | 41.0 s | 39.3 s ^e |
| 23 | =CH | 6.10 dd | 16.0 | 144.2 d | 6.00 dd | 16.0 | 143.6 d | 144.1 d ^e |
| | | | 10.0 | | | 10.0 | | |
| 24 | =CH ₂ | 5.04 dd | 16.0 | 114.3 t | 5.20 dd | 16.0 | 114.3 t | 113.1 t ^e |
| | | | 2.0 | | | 2.0 | | |
| | | 5.07 dd | 10.0 | | 5.25 dd | 10.0 | | |
| | | | 2.0 | | | 2.0 | | |
| 25 | CH ₃ | 1.08 s | - | 22.6 q | 1.02 s | - | 22.6 q | 27.4 q ^e |
| 26 | CH ₃ | 1.16 s | - | 22.9 q | 1.16 s | - | 23.0 q | 27.4 q ^e |
| 27 | C=O | - | - | - | - | - | 171.7 s | - |
| 28 | CH ₃ | - | - | - | 2.59 s | - | 24.2 q | - |

^aMeasured in CDCl₃ solution. ^bCalculated for *o*-toluidine. See reference 9).^cCalculated for *o*-tolylisocyanate. See reference 9). ^eObtained from reference 4).

Furthermore, the validity of this structure was proved by acetylation of 1. Treatment of 1 with acetic anhydride in pyridine at reflux for 3 hours gave mono- and di-acetates (2 and 3). Monoacetate of 1, mp. 242-244°C, was identified as LL-S4908 in comparison with IR, UV, ¹H-NMR and MS spectra. The structure of diacetate [δ 1.38 (3H, s, CH₃CO-), 2.68 (3H, s, CH₃CO-)] was determined as 3¹⁰ in which -NH-s of indoline and benzodiazepine moiety were acetylated. On acetylation with the same condition, LL-S4908 also gave 3.

Application of 1 at a concentration of 50 µg/ml apparently induced the abnormal second cleavage of the sea urchin embryos.

References and Footnotes

- 1) G. A. Ellestad, P. Mirando and M. P. Kunstmann, J. Org. Chem., 24, 4204 (1973).
- 2) 1: *Anal.* Found :C, 73.68;H, 6.33;N, 11.28, Calcd. for C₂₃H₂₃N₃O₂:C, 73.97; H, 6.21;N, 11.25;O, 8.57 %.
- 3) A. J. Birch and J. J. Wright, Tetrahedron, 26, 2329(1970).
- 4) H. Nagasawa, A. Isogai, A. Suzuki and S. Tamura, Agric. Biol. Chem., 43 1759(1979).
- 5) R. Marchelli, A. Dossena, A. Pochini and E. Dradi, J. Chem. Soc. Perkin I, 713(1977).
- 6) J. H. Beynon, R. A. Sanders and A. E. Williams, "The Mass Spectra of Organic Molecules", Elsevier, New York, N. Y., 1968, p.303.
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- 8) P. K. Martin, H. Rapoport, H. W. Smith and J. L. Wong, J. Org. Chem., 34, 1359(1969).
- 9) G. C. Levy and G. L. Nelson, "Carbon-13 Nuclear Magnetic Resonance for Organic Chemist", Wiley-Interscience, a Division of John Wiley & Sons, Inc., New York, N. Y., 1972, p.81.
- 10) 3: White amorphous mp. 218-221°C; UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm(ϵ): 206 (72,200), 237 (25,400), 277 (5000); IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 1735, 1720, 1688, 1650,1600; ¹H-NMR (in CDCl₃): 0.90 (3H, s), 1.12 (3H, s), 1.38 (3H, s), 2.68 (3H, s), 2.54 (1H, dd, J=14.0, 8.5 Hz), 3.12 (1H, d, J=14.0 Hz), 4.22 (1H, d, J=8.5 Hz), 5.08 (1H, dd, J=16.0, 2.0 Hz), 5.13 (1H, dd, J=10.0, 2.0 Hz), 5.86 (1H, dd, J=16.0, 10.0 Hz), 6.11 (1H, s), 7.02-8.02 (m, 8 x Ar-H); MS m/z: 457 (M⁺), 415 (M⁺-42), 346 (M⁺-42-69), 304 (M⁺-42-69-42), 130.

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